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Isotope-induced Nonequivalence in a Symmetrical Molecule: Measurement of the ³¹P-³¹P Geminal Coupling Constant in Pyrophosphate

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Asymmetric incorporation of ¹⁸O into pyrophosphate to perturb the ³¹P n.m.r. resonance frequency allows determination of the ³¹P–³¹P geminal coupling constant in this otherwise symmetrical molecule.

We report the use of selective ¹⁸O labelling of one phosphorus atom in pyrophosphate to induce chemical shift nonequivalence of the two phosphorus nuclei and thus permit observation of the individual resonances and measurement of an otherwise unobservable mutual coupling constant.

¹⁸O induces a small upfield shift in the ³¹P n.m.r. resonance frequency of phosphates (*ca.* 0.02 p.p.m./¹⁸O bond), the magnitude of which is related to the P–O bond order.^{1–3} Recent work in this laboratory involving the synthesis of ¹⁸O- β , γ -bridge-labelled ATP for use in positional isotope exchange experiments produced an interesting result while assigning resonances in the ³¹P n.m.r. spectra of ¹⁸O-labelled pyrophosphate intermediates.⁴ At 202.5 MHz, the ³¹P n.m.r. spectrum of pyrophosphate with one nonbridging ¹⁸O appeared as a closely spaced doublet with a separation of 0.5 Hz. This splitting was presumed to be due to coupling between the nonequivalent phosphorus atoms with the observed doublet corresponding to the central doublet of an AB pattern. By selectively increasing the number of ¹⁸O atoms attached to one of the two phosphorus atoms in the pyrophosphate molecule we can increase the nonequivalence of the phosphorus atoms sufficiently to allow observation of



Figure 1. (a) Simulated ³¹P n.m.r. spectrum of ¹⁸O₄- and ¹⁸O₃labelled pyrophosphate generated by summing weighted subspectra of the illustrated compounds. Simulations were done on a Nicolet 1180 computer using the Nicolet program NTCSIM. Linewidths were set to 1.5 Hz and a coupling constant of 21.1 Hz was used in all simulations. Chemical shifts were calculated as described in the text. (b) ³¹P N.m.r. spectrum at 202.5 MHz of $[\alpha$ -¹⁸O₄]pyrophosphate contaminated with 13% $[\alpha$ -¹⁸O₃]pyrophosphate. The n.m.r. sample was prepared by dissolving the tetrasodium salt (19 mg) in 0.4 ml of 50% D₂O containing 0.5 mM ethylene glycol bis(β -aminoethyl ether)-N,N,N',N'-tetraacetic acid. A 5 mm acid-washed n.m.r. tube was used. The spectrum was obtained using a Bruker 500 MHz spectrometer located at the Southern California Regional NMR Facility, California Institute of Technology. The sample was spun at 17 \pm 1 Hz and 128 accumulations were made using a spectral width of 2000 Hz. The free induction decay was collected using 8192 data points followed by zero-filling to 16 K before Fourier transformation.

the outer resonances of the AB pattern. From this pattern the ${}^{31}P{}^{-31}P$ geminal coupling constant in pyrophosphate can be easily determined.

 $[\alpha$ -¹⁸O₄]Pyrophosphate was prepared from $[\gamma$ -¹⁸O₄]ATP using yeast acetyl coenzyme A synthetase.^{5,6} $[\gamma$ -¹⁸O₄]ATP was prepared by the method of Hoard and Ott⁷ from ADP and $[^{18}O_4]$ phosphate [prepared, in turn, from H₂¹⁸O (99%) and PCl₅].⁶ The isotopic purity of the $[^{18}O_4]$ phosphate was determined by mass spectral analysis of the trimethyl ester derivative, which was prepared by methylation of the free acid with diazomethane in diethyl ether.⁹⁻¹¹

Figure 1(b) shows the 202.5 MHz spectrum of α -¹⁸O₄labelled pyrophosphate. The ³¹P-³¹P geminal coupling constant in this AB pattern is simply the separation between either the two downfield or the two upfield peaks and was measured to be 21.1 Hz, a value comparable to the ³¹P-³¹P coupling constants found in ATP.¹² The uncoupled chemical

shift difference of the two phosphorus resonances was calculated to be 13.3 Hz (at 202.5 MHz) for our labelled pyrophosphate.13 Based on a net difference of four P-18O bonds, the observed chemical shift difference gives an upfield shift due to ¹⁸O of 0.0164 p.p.m./bond, which is consistent with published values.1 The unequal peak heights of the central doublet and the shoulder appearing on the downfield side of the central doublet were attributed to the presence of contaminating $[\alpha^{-18}O_3]$ pyrophosphate. Figure 1(a) shows the simulated spectrum generated from three separately simulated subspectra which were weighted appropriately and then summed. Chemical shifts of the pyrophosphates were calculated based on the experimentally determined upfield shift of 0.0164 p.p.m./18O bond. In the case of 18Obridged $\left[\alpha^{-18}O_{3}\right]$ pyrophosphate the chemical shift difference used in the simulation was calculated using 3.67 ¹⁸O bonds to the α -phosphorus (an average of the three resonance forms).

Since scalar coupling constants between equivalent nuclei cannot be determined using magnetic resonance experiments,^{14,15} the magnetic environment of one or more of the nuclei must be altered, usually by chemically modifying the molecule. The use of isotope-induced chemical shift perturbations as illustrated in this work permits the measurement of coupling constants between equivalent nuclei without modification of either structure or chemistry. Oxygen isotope-effects (specifically, ¹⁶O and ¹⁸O) are especially useful in this respect since the observed isotope-effect is due solely to a change in mass. Complications arising from changes in the spin quantum number (giving rise to additional coupling) can therefore be avoided.

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References

- 1 M. Cohn, Annu. Rev. Biophys. Bioeng., 1982, 11, 23.
- 2 M. Tsai, Methods Enzymol., 1982, 87, 235.
- 3 M. Cohn and A. Hu, J. Am. Chem. Soc., 1980, 102, 913.
- 4 M. A. Reynolds, N. J. Oppenheimer, and G. L. Kenyon, J. Am. Chem. Soc., in the press.
- 5 P. Berg, J. Biol. Chem., 1956, 222, 991.
- 6 P. D. Boyer, O. J. Koeppe, and W. W. Luchsinger, J. Am. Chem. Soc., 1956, 78, 356.
- 7 D. E. Hoard and D. G. Ott, J. Am. Chem. Soc., 1965, 87, 1785.
- 8 J. M. Risley and R. L. Van Etten, J. Labelled Comp. Radiopharm., 1978, 15, 533.
- 9 C. F. Midelfort and I. A. Rose, J. Biol. Chem., 1976, 251, 5881.
- 10 M. J. Wimmer and I. A. Rose, J. Biol. Chem., 1977, 252, 6769.
- 11 D. H. Eargle, V. Licko and G. L. Kenyon, Anal. Biochem., 1977, 81, 186.
- 12 M. Cohn and T. R. Hughes, J. Biol. Chem., 1960, 235, 3250.
- 13 E. D. Becker, 'High Resolution NMR,' Academic Press, New York, 1980, pp. 135-139.
- 14 H. S. Gutowsky, D. W. McCall, and C. P. Slichter, J. Chem. Phys., 1953, 21, 279.
- 15 A. Abragam, 'The Principles of Nuclear Magnetism,' Oxford University Press, London, 1961, pp. 480-495.